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COMPARATIVE ANALYSIS OF GENE EXPRESSION PROFILES AMONG CONTRASTING MULBERRY VARIETIES UNDER COLD STRESS CONDITION

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ABSTRACT

Low temperature/ cold stress is a major abiotic stress affecting mulberry plant growth and productivity in temperate climatic conditions. Mulberry leaf is a sole food source for the economically important insect, *Bombyx mori*. Cold stress severely damages buds and young leaves and affecting growth and leaf productivity. Therefore, it is required to identify genes which are responsive and impart cold tolerance and utilize them to develop improved mulberry varieties, which can withstand cold stress without compromising its leaf production. In the present study, candidate cold tolerant gene sequences of WRKY46, Spermidine synthase, early response to dehydration10 (ERD10), TIFY10, Cysteine proteinase Inhibitor (CPI), Cold regulated 413 plasma membrane protein 2 (COR413) were retrieved from the mulberry database through NCBI and bioinformatics tools. Comparative gene expression analysis was performed in temperate mulberry varieties, Gosherami, Leh 2 (saspole) and Gurez using tropical variety, G4 as a reference in cold stress condition. Results depicted that WRKY46, Spermidine synthase, ERD10, TIFY10, CPI, COR413 genes showed significant up-regulation under cold stress condition. Gurez genotype displayed significant up-regulation of these selected cold tolerant genes under both laboratory and field conditions compared to other mulberry genotypes.

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1 Introduction

Plants grown under temperate climatic condition faced freezing or extreme low temperature condition. In general, plants from temperate climate are found to be chilling tolerant with variable degree, and can increase their freezing tolerance by being exposed to chilling, non-freezing temperatures. These temperate plants have evolved a mechanism via cold acclimation to enhance tolerance to low temperatures (Thomashow, 1999). In addition, the same mechanism used by perennial woody plant species of temperate regions to tolerate freezing (Levitt, 1980; Sakai & Larcher, 1987). However, the high freezing tolerance exhibited by perennial woody plants is regulated in response to cues such as the short day and decreased air temperatures during autumn (Sakai & Larcher, 1987). In many herbaceous and perennial woody plants, including mulberry, freezing tolerance is dependent on occurrence of numerous biochemical and physiological changes (Sakai & Yoshida, 1968; Yoshida, 1984; Fujikawa, 1994). These changes brought at cellular level through compositional changes in the plasma membranes (Steponkus, 1984; Yoshida, 1984; Uemura et al., 1995), enhanced accumulation of compatible osmolytes, such as soluble sugars, prolines, and betaines (Hare et al., 1998), induction of gene expression such as heat shock proteins (Neven et al., 1992; Ukaji et al., 1999), cold-regulated (COR) proteins (Guy et al., 1985; Thomashow, 1999), increased extracellular accumulation of antifreeze proteins (Griffith & Antikainen, 1996), and changes in the property of cell walls (Rajashekar & Lafta, 1996; Fujikawa and Kuroda, 2000) were found to be associated with freezing tolerance (Guy, 1990; Fujikawa et al., 1999; Thomashow, 1999).

Genes such as WRKY46, Spermidine synthase, early response to dehydration10 (ERD10), TIFY10, Cysteine proteinase Inhibitor (CPI), cold regulated 413 plasma membrane protein 2 (COR413) have been shown to be involved in cold stress acclimation in different model and crop plants (Martin-Tanguy, 2001; Breton et al., 2003; Puhakainen et al., 2004; Wang et al., 2012; Huang et al., 2016). WRKYs are transcription factors which regulate numerous downstream genes involved in stress adaptation by binding to W-box sequences present in the promoter region. WRKY genes were reported to be involved in different stresses and shown to be upregulated during cold stress in mulberry (Wang et al., 2012; Ding et al., 2014; Baranwal et al., 2016). Dehydrin belongs to the group 2 LEA (Late Embryogenesis Abundant II) proteins. These proteins accumulate in late embryogenesis or can be induced in response to dehydration, salt, cold and freezing stress (Wisniewski et al., 1999; Puhakainen et al., 2004; Rorat et al., 2006; Hanin et al., 2011). Spermidine synthase is a key enzyme in polyamine spermidine biosynthesis (Kasukabe et al., 2004). Polyamine spermidines are low molecular weight aliphatic amines and play

an important role in growth, development reproduction and stress adaptation in plants (Kakkar et al., 2000; Martin-Tanguy, 2001).

The TIFY are novel plant specific genes are characterized by a conserved TIFY motif (TIFF/YXG) (Zhu et al., 2014). TIFY genes were reported to involve in jasmonate signaling, plant growth, development, pathogen response and in response to abiotic stress (Zhu et al., 2011; Hakata et al., 2012; Zhu et al., 2012). Upregulation of *TIFY* genes under cold stress was reported in moso bamboo (*Phyllostachys edulis*) (Huang et al., 2016) and rice (Ye et al., 2009). Cysteine proteinase inhibitors, small peptides of 12-16 kDa, are proteinaceous inhibitor of papain (Benchabane et al., 2010). Reports suggests that cysteine proteinase inhibitor play an important role in plants exposed to different stresses including cold stress (Zhang et al., 2008; Benchabane et al., 2010; Kunert et al., 2015; Subburaj et al., 2017).

Cold regulated 413 plasma membrane protein 2 belongs to COR413 family and having five putative transmembrane domain (Breton et al., 2003). Cold regulated 413 plasma membrane protein 2 (COR413-PM2) was first reported in wheat and Arabidopsis and accumulation of its transcript correlated with freezing tolerance in these plants (Breton et al., 2003).

Sericulture is practiced in temperate climatic conditions where, mulberry, the sole food source for the silkworm, *Bombyx mori* encounters cold stress (Rohela et al., 2016; Rohela et al., 2018a; Rohela et al., 2018b; Shabnam et al., 2018). Due to cold/frost, upper branch of the mulberry plant get damage which in turn affect the leaf yield for the spring rearing of silkworm (Shukla et al. 2016a). To the extent of 30% frost damage of mulberry plant was reported during temperate climate of Kashmir valley, India (Ahanger et al., 2013). This is due to existing genotypes used in sericulture industry are not frost tolerant. However few genotypes from Leh, Ladakh and Gurez regions, India displayed high degree of cold adaptation, but they are not commercially exploited. The molecular mechanism of strong adaptability and tolerance to cold stress has not been studied for these genotypes, which limits its exploitation. Therefore, it is required to understand the genes involved in cold tolerance mechanism in these mulberry genotypes to utilize in mulberry breeding.

In present study, a comparative gene expression study of WRKY46, Spermidine synthase, Early response to dehydration10 (ERD10), TIFY10, Cysteine proteinase Inhibitor (CPI), Cold regulated 413 plasma membrane protein 2 (COR413) was performed in temperate mulberry, Gosherami, Leh 2 (Saspole) and Gurez along with tropical mulberry germplasm G4 and V1 in cold stress condition. Present study was carried out with an aim to observe the expression pattern of these genes under cold stress condition for their utilization in mulberry improvement.

2 Materials and Methods

2.1 Retrieval of gene sequences, Multiple Sequence Alignment, Phylogenetic analysis and Analysis of gene structure

Putative cold tolerant gene sequences were retrieved from the Mulberry genome database MorusDB and comparative sequences analysis using NCBI-BLAST. Similar, cold tolerant gene sequences were also retrieved from other plant species. These sequences were arranged in fasta format and multiple sequence alignment was performed using MUSCLE (<https://www.ebi.ac.uk/Tools/msa/muscle/>). Based on the alignment, the phylogenetic tree was constructed from the amino acid sequences using the neighbor-joining (NJ) method in MEGA 7 with 1000 bootstrap replicates. The exon-intron structures of the gene sequences were determined using Gene Structure Display Server (GSDS:<http://gsds.cbi.pku.edu.cn>) by aligning cDNA to their corresponding genomic DNA sequences.

2.2 Plant materials and cold stress treatments

Mulberry (*Morus alba*) variety G4 grown in nursery and transplanted to pots. Similarly, Gosherami, Leh 2 (Saspole) and Gurez variety were grown, buds from these varieties were grafted on to G4. These bud grafted cuttings were grown under well watering condition in plastic pots for three months. For cold treatments, plants were transferred to incubator at 4°C and samples were collected at 6 hrs, 24 hrs and 48 hrs.

2.3 RNA extraction and cDNA synthesis

Total RNA extraction was performed as per the manufacturer's

instructions using NucoSpin plant RNA extraction kit (TaKaRa, Japan). Quantification of RNA samples were determined using the NanoDrop 2000c spectrophotometer (Thermo Fisher Scientific, USA) and the quality of RNA samples was analyzed on ethidium bromide stained-agarose gels. First-strand cDNA synthesis was performed according to manufacturer's instructions through the PrimeScript 1st strand cDNA synthesis kit (TaKaRa, Japan). Further, 10 fold dilution of cDNA was carried out with nuclease free sterile water for qRT-PCR (Shukla et al., 2016a; Ahmed et al., 2018).

2.4 Primer design

Primers for expression study were performed using Primer-Blast software (www.ncbi.nlm.nih.gov/tools/primer-blast) with product size of 100-180 bp and primer length 20-25 bp. The details of the primers used in this study are given in Table 1.

2.5 Quantitative Real Time PCR

Quantitative real time PCR (qPCR) was performed using SYBR® Premix Ex Taq™ II (Tli RNaseH Plus) (TaKaRa, Japan) on Stratagene Mx3005P (Agilent Technologies). A reaction mixture of 10 µl was prepared having cDNA template (1 µl), primer (0.2mM forward & reverse each), and SYBR Premix Ex Taq II (1X). The PCR was set in a 96-well reaction plate at 94°C for 30 s, followed by 40 cycles of 94°C for 10 s, 55°C for 30 s, and 72°C for 30 s. After 40 cycles, the melting curve was determined to verify primer specificity by heating from 65°C to 95°C. Each qPCR reaction was performed in two technical replicates and three biological replicates (Ahmed et al. 2018).

Table 1 Details of primers sequences used for the study

S. No.	Name	Sequence (5' - 3')
1.	WRKY46 qRT Forward	GAGGTCACTTACAGAGGCCG
2.	WRKY46 qRT Reverse	GACCTGTTGCTTGTTGGTTGC
3.	ERD10 qRT Forward	CTATCGCCGGTGACTTCGAG
4.	ERD10 qRT Reverse	AACCGGGCTTTTCTCTCTCC
5.	TIFY10 qRT Forward	CGGTTCTTGGAGAAGAGAAAG
6.	TIFY10 qRT Reverse	TCATTGGGTTTGTTGAGCAGC
7.	Spermidine qRT Forward	AAGTTGCTCGGCATACCTCC
8.	Spermidine qRT Reverse	GGGACAGCCTTCAAAAAGGC
9.	COR413 qRT Forward	CAAGCTGAAAACCAGGGCAC
10.	COR413 qRT Reverse	CCCTCCCCATTCAGCTCTC
11.	CPI qRT Forward	GAAACTCGGCGGAATCCAAG
12.	CPI qRT Reverse	AGACCGTACCGGCAACAAC

3 Results

3.1 Identification of candidate cold tolerant gene sequences from mulberry

WRKY46, Spermidine synthase, CPI, ERD10, COR413, TIFY10 gene sequences were retrieved from MorusDB using NCBI-BLAST. *M. notabilis* used as a reference for identification of candidate cold tolerant genes from mulberry as whole genome sequence and annotation is publicly available. Nucleotide and amino acid sequence of *M. notabilis* WRKY46 was 1095 bp and 364 aa respectively (accession no. XM_010097487.1). Similar gene sequence was also identified from *M. alba* (accession no. KP765436.1), which exhibited 96.99% nucleotide and 94.51% amino acid identity with *M. notabilis* sequence. Spermidine synthase gene from *M. notabilis* codes for 1005 bp nucleotide sequence and 334 aa (accession no. XM_010102183.1). Similar, but partial sequence of spermidine synthase was also identified from *M. nigra* (accession no. KP980553.1). Cysteine proteinase inhibitor of 306 bp and 101 aa was also fished out from *M. notabilis* (accession no. XM_010108020.1) and *M. alba* (accession no. KM091230.1) that exhibited 99.35% and 99.01% nucleotide and amino acid identity respectively. ERD10 gene was identified as 759 bp and 252 aa. *M. notabilis* COR413 gene sequence was identified having 606 bp nucleotide and 201 aa sequence. TIFY10A contained 822 bp and 273 aa (accession no. XM_010100794.1) of nucleotide and amino acid sequence respectively. EST sequence showing identity with TIFY10A (ES448372.1), WRKY46 (ES448888.1), spermidine synthase (JK705754.1), cysteine proteinase inhibitor (ES449068.1) were also retrieved from the EST database of mature leaf of *M. indica*.

3.2 Multiple sequence alignment, Phylogenetic analysis, and gene structure of cold tolerant genes

Multiple sequence alignment of WRKY46 of different plant species revealed that *M. notabilis* WRKY46 shows 55.21%, 41.52%, 39.52%, 30.59%, 28.57%, 27.85% similarity with *Populus trichocarpa*, *Brassica rapa*, *Arabidopsis thaliana*, *Gossypium hirsutum*, *Zea mays*, *Triticum aestivum* respectively and having conserved WRKY specific domain. Phylogenetic analysis also showed that *M. notabilis* displayed highest similarity with *M. alba* and *Populus trichocarpa*. Alignment of *M. notabilis* CPI revealed the presence of inhibitory domain and showed overall similarity of 99.01%, 70.10%, 69.70%, 66.00%, 63.00% with *M. alba*, *Coffea canephora*, *Boehmeria nivea*, *Cajanus cajan*, *Hevea brasiliensis*, respectively. Similar results were observed with phylogenetic analysis. Sequence alignment and phylogenetic analysis of *M. notabilis* ERD10 showed highest similarity with *Hevea brasiliensis* and presence S-segment (SSSSSS) and K-segment (EKIKEKLP) specific to ERD10. Sequence alignment of TIFY 10A showed the presence of TIFY motif (TIFF/YXG) and Jas domain. Sequence alignment and phylogenetic analysis of *M. notabilis* TIFY10A with different plant species showed highest similarity with *Malus domestica*. Multiple sequence alignment and phylogenetic analysis of *Morus* spermidine synthase gene sequences with different species shows its close similarity with *Populus trichocarpa* (86.03%) and *Malus domestica* (84.76%). COR413 gene sequence alignment with other species shows close identity with *Hevea brasiliensis* (88.56%) and *Malus domestica* (84.08%). Phylogenetic analysis of COR413 also revealed the same result. The gene structure features like exon-intron distribution and conserved motif of *M. notabilis* WRKY46, Spermidine synthase, Cysteine proteinase inhibitor, ERD10, COR413, TIFY10 gene sequences were analyzed. ERD10 and cysteine proteinase inhibitor contain 02 exons, WRKY46 contain 3 exons, COR413 contain 04 exons, TIFY10A with 05 exons, spermidine synthase contain 09 exons (**Figure 1**).

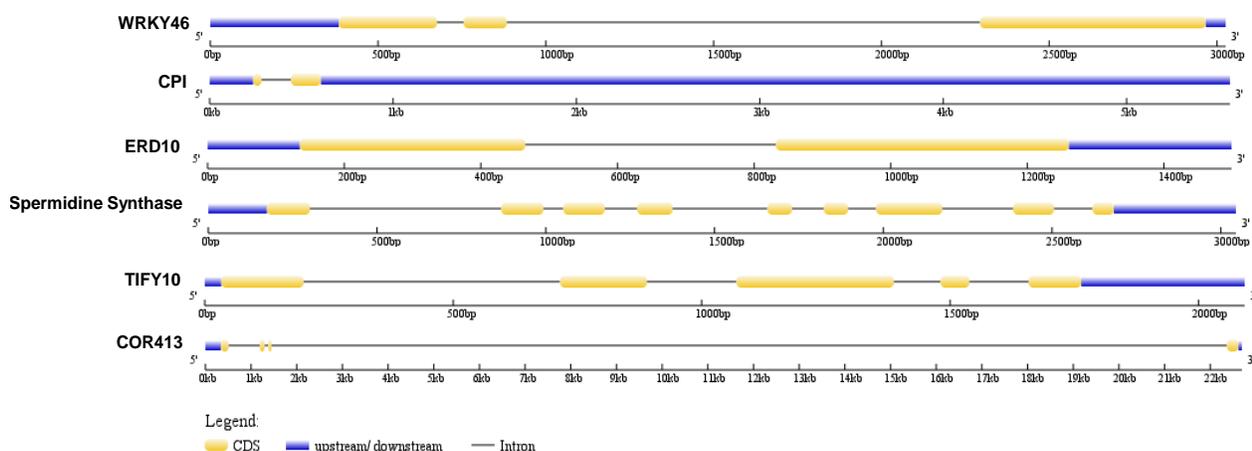


Figure 1 Gene Structure of WRKY46, CPI, ERD10, Spermidine synthase, TIFY10, COR413.

3.3 Comparative expression profile of candidate genes in cold stress condition:

Quantitative PCR was performed for the *in silico* identified cold tolerant genes and relative expression analysis was carried out in tropical (G4 genotype) and temperate mulberry genotypes (Gosherami, Leh 2 (Saspol) and Gurez) upon exposure to cold stress. Comparative expression profile performed by two methods, where control samples without stress from the respective genotypes used as a reference and in other method, expression levels of tropical genotype, G4 as a reference sample. It was observed that *in silico* identified putative cold tolerant genes like WRKY46, Spermidine synthase, ERD1, TIFY10, CPI, COR413 displayed upregulation during cold stress condition. WRKY46 gene displayed nearly 31 fold upregulation in Gosherami, leh 2 and gurez genotype while 6 fold in G4 genotype compared to untreated samples (**Figure 2 A**). But, WRKY46 gene showed 5.8 fold upregulation in gurez genotype, 2.4 fold upregulation in Gosherami and 1.8 fold upregulation in Leh 2 genotype when G4 expression used as a control (**Figure 2G**). Under cold stress condition, Spermidine synthase was found to be upregulated 16.7 fold in Gurez, 15.7 fold in Gosherami, 3.85 fold in G4 and 2.74 fold in Leh 2 compared to untreated sample as a control in contrast to G4 used as a control (**Figure 2B**). Spermidine synthase expression was downregulated in Gosherami and Leh 2 while 5.7 fold upregulated in Gurez (**Figure 2H**). ERD10 expression was 129 fold in Gosherami, 24 fold in Gurez, 2.5 fold in G4 and down regulated in Leh2 genotype compare to untreated sample as a control (**Figure 2C**). The expression of ERD10 induced to 351 fold in Gosherami, 7.4 fold in Gurez and 5.6 fold in Leh 2 when G4 used as a control (**Figure 2I**). TIFY10 gene expression was highest in Gurez 144.76 fold followed by Gosherami 102 fold and in G4 10.87 fold and Leh 2, with 5.7 fold when untreated sample used as control (**Figure 2D**). When G4 used as a control, Gurez displayed highest expression of 6.7 fold while Gosherami and Leh2 showed 2.9 and 2.0 fold respectively (**Figure 2J**). CPI expression compared to untreated control was 9.66 fold in Gosherami, 8.77 fold in Gurez, 4.71 fold in leh 2 and 1.86 fold in G4 genotype (**Figure 2E**). When data was compared with G4 as a control, Gurez showed highest expression of 9.6 fold, followed by Leh 2 with 4 fold and Gosherami with 2.8 fold (**Figure 2K**). COR413 expression was observed highest 12 fold in Gurez, 3 fold in Gosherami and 2 fold in Leh 2 and G4 under cold stress condition when untreated sample used as a control (**Figure 2F**) but when G4 used as a control only gurez displayed 9.5 fold expression of COR413 while downregulation of COR413 expression was observed in Gosherami and Leh 2 (**Figure 2L**).

3.4 Comparative expression profile of candidate cold tolerant genes under field conditions

Laboratory and controlled conditions experiment results suggest that Gurez genotype exhibited highest up-regulation of most of the tested cold tolerant genes compared to Gosherami and Leh2 genotype. To further validate the expression profile in Gurez, RNA was isolated from the sample collected from the tropical mulberry variety, V1, and temperate varieties, Narula and Gurez under field cold stress condition ($6^{\circ}\text{C} \pm 2$) from the mulberry germplasm bank at CSR&TI, Pampore, India. The relative expression of selected genes was analyzed in these samples in which V1 was used as reference for normalizing the relative expression of genes. Upregulation of WRKY46, Spermidinesynthase, CPI, ERD1, COR413, TIFY10 was observed in Gurez genotype compare to Narula genotype. Expression of WRKY46 was 22 fold in gurez while 3 fold in Narula, Spermidine synthase expression was 7 fold in gurez while 4 fold in narula, CPI expression was 15.5 fold in gurez while 4 fold in narula, ERD10 expression was 4.6 fold in gurez while 2 fold in narula, COR413 expression 9 fold in gurez while 7 fold in narula, TIFY10 expression 13.5 fold in gurez while 5.7 fold in narula (**Figure 3**).

4 Discussion and conclusions:

Freezing or extremely low temperature constitutes a key factor in temperate region by influencing plant growth, development and productivity. Plants in temperate region of Kashmir affected by cold stress especially mulberry plants, which is important for sericulture Industry (Shukla et al., 2016b). Kashmir valley is famous for its bivoltine temperate sericulture. However, being a temperate region, it is important to mention that the mulberry genotypes used for sericulture were affected by frost/cold damage. However, there are genotypes identified from Leh, ladakh and gurez region that displayed high degree of tolerance to cold. Understanding the mechanism of tolerance and genes involved in cold tolerance can be useful for development of cold tolerant varieties. Mulberry genome (*M. notabilis*) was sequenced in 2013 and in addition, large amount of transcriptomic data of *M. notabilis* (a haploid genome $n=7$) was generated (He et al., 2013). These data are increasingly used for comparative genomics, identification of many useful genes and analysis of non-coding sequences (He et al., 2013; Ma et al., 2014; Wang et al., 2014; Wei et al., 2014; Song et al., 2016;). Recently, Transcriptomic data from *M. laevigata* and *M. serrata* was generated (Saeed et al., 2016) and utilized for identification and genome wide analysis of WRKY and NAC transcription factor (Baranwal & Khurana, 2016; Baranwal et al., 2016).

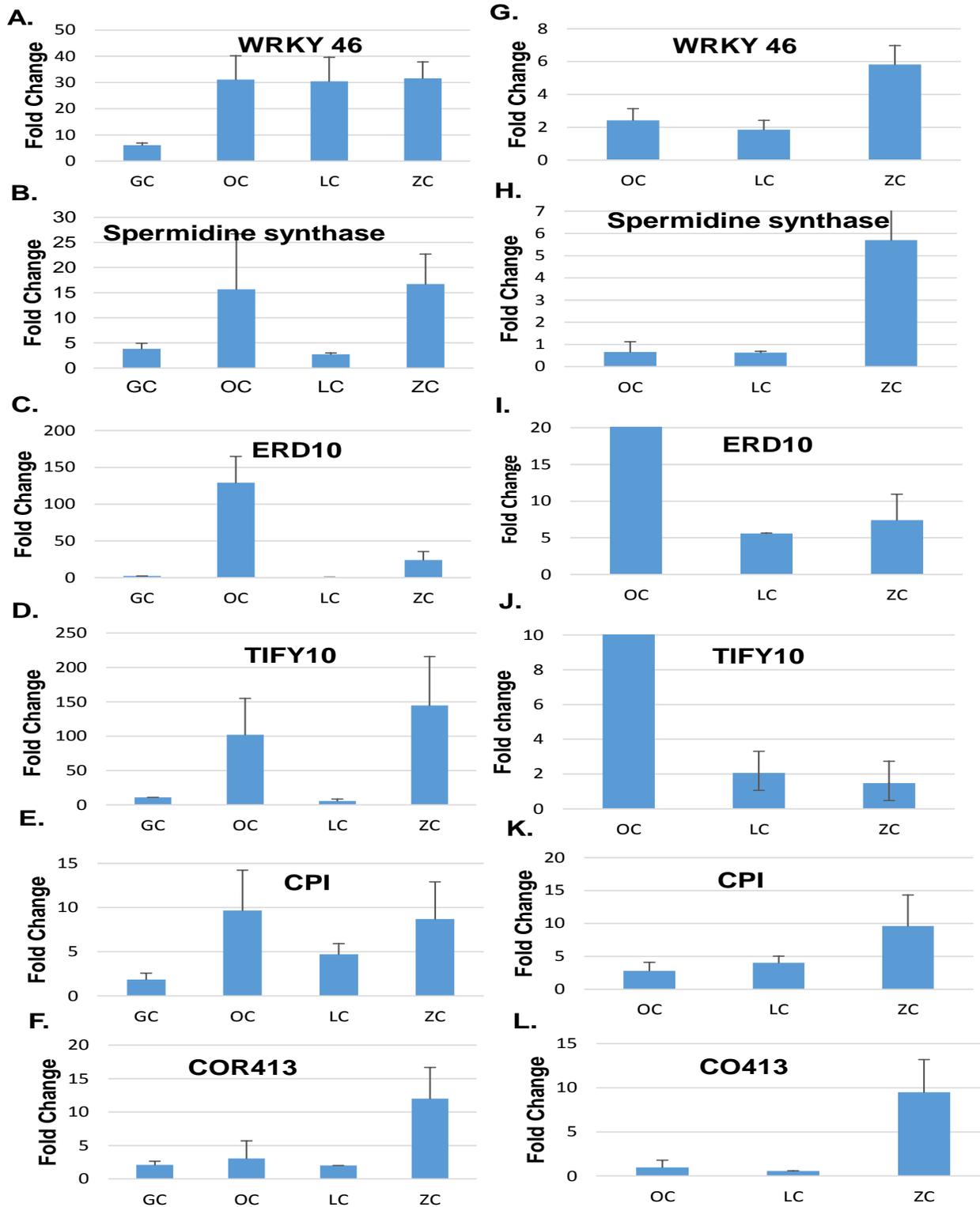


Figure 2: Relative expression of WRKY46, Spermidine synthase, ERD10, TIFY10, CPI, COR413 under cold stress under laboratory conditions. The relative expression of genes in cold stress as compared un-treated samples as control (A, B, C, D, E & F). The relative gene expression of genes compared to cold stress treated G4 samples as control (G, H, I, J, K & L). GC: G4; OC: Goshalami; LC: Leh 2; ZC: Gurez.

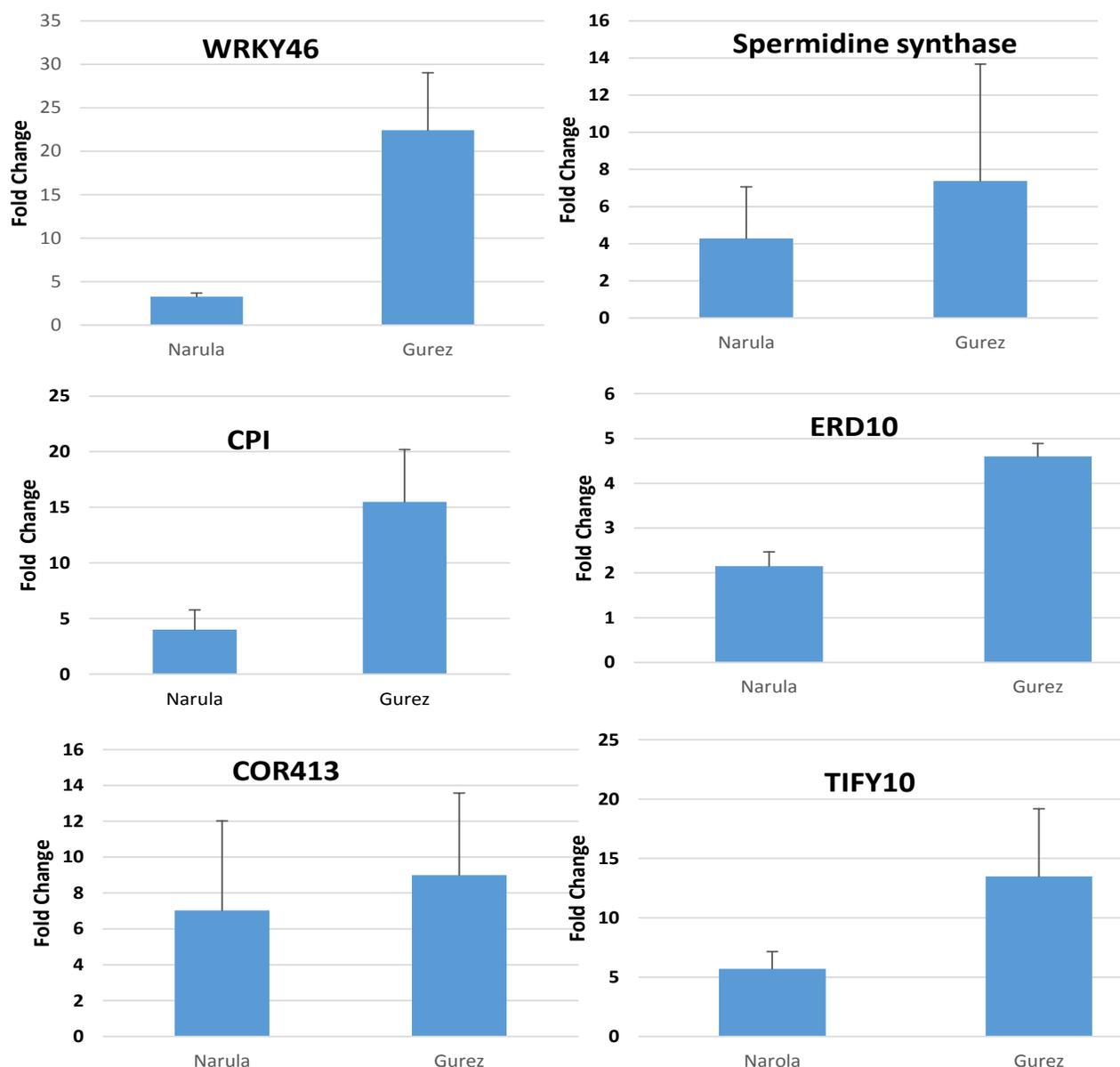


Figure 3 Relative expression of WRKY46, Spermidine synthase, ERD10, TIFY10, CPI, COR413 under cold stress from field conditions. Leaf samples of narula and gurez were collected from Germplasm Bank of CSR&TI, Pampore under low temperature condition (Temperature: $6^{\circ}\text{C} \pm 2$).

Results of present study indicated the up-regulation of WRKY46, ERD10, spermidine synthase, TIFY10, Cysteine proteinase inhibitor, and COR413 when exposed to cold stress condition both in laboratory and field conditions. Many reports also showed the up-regulation of these genes under cold stress condition in different crop plants. For instance, *Brassica compestris* WRKY46 (*bcWRKY46*) was previously reported to enhance cold tolerance in transgenic tobacco. Baranwal et al. (2016) reported the differential

regulation of mulberry WRKY genes under different abiotic stress conditions including cold stress. Dehydrins like DHN, ERD10, LEA etc. were reported to be enhance freezing tolerance in *Arabidopsis* and also involve in different abiotic stress tolerance (Puhakainen et al., 2004; Rorat et al., 2006; Hanin et al., 2011). Spermidine synthase was found to involve in plant growth and development (Kakkar et al., 2000; Martin-Tanguy, 2001) and also provide chilling tolerance in cucumber cultivar (Shen et al., 2000).

Previous study reported the involvement of TIFY 10 in alkaline stress (Zhu et al., 2014), while in present study up-regulation of TIFY10 expression during cold stress in different mulberry genotypes was reported. This indicated that this gene might also involve in cold stress tolerance. Cysteine proteinase inhibitor (CPI) genes are expressed in plants exposed to high salt, drought, cold and heat stress (Zhang et al., 2008; Je et al., 2014; Kunert et al., 2015). In *Arabidopsis*, expression of cysteine proteinase inhibitor conferred the cold tolerance in transgenic plants (Zhang et al., 2008). Cold regulated 413 plasma membrane protein 2 transcript accumulation was correlated with freezing tolerance in *Arabidopsis* and wheat (Breton et al., 2003). In *Brachypodium*, accumulation of *BradiCOR413* transcripts was reported after cold treatment (Colton-Gagnon et al., 2013). Over expression of tomato *SICOR413IMI* improved chilling tolerance in transgenic plants (Ma et al., 2018).

Comparison of candidate cold tolerant gene expression profile between genotypes found that temperate genotypes showed higher gene expression levels compared to tropical genotypes. Among the temperate genotypes, gurez genotype displayed the highest level of cold tolerant gene expression. These results suggest that there is a genetic and molecular difference between genotypes for acclimation to cold stress resulting from differential expression and regulation of cold tolerant genes.

In conclusion, the analyzed candidate genes are induced in response to cold stress and among the genotypes Gurez displayed highest upregulation under both laboratory and field conditions suggests these genes might involve in cold tolerance in this genotype. Further, these genes from Gurez genotype are good candidate for development of cold tolerant mulberry varieties.

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