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STUDYING INDICATORS OF GELATIN AND NATURAL POLYSACCHARIDES QUALITY AND SAFETY

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ABSTRACT

Present study was conducted with the aim of finding out the optimal composition of biodegradable packaging materials based on gelatin and natural polysaccharides, film samples. The film samples have been obtained with the purpose of choosing the optimum composition of biodegradable packaging materials based on gelatin and natural polysaccharides. Physico-chemical properties and safety performance of gelatin and natural polysaccharides (agar, carrageenan, hydroxypropylmethicellulose – HPMC) have been studied. Films based on gelatin and natural polysaccharides have been obtained by drying solutions of the tested components at various temperatures. Result of study revealed the fact that physico-chemical parameters of the studied samples of agar-agar and hydroxypropymethylcellulose (HPMC) confirmed to the requirements of normative documents. In terms of chemical and microbiological safety, both studied samples of agar-agar and carageenan satisfy the applicable regulatory requirements of the international food standard, and may be used as raw material for obtaining biodegradable polymers. It has been proved that the studied samples of carageenan meet the requirements of applicable normative documentation in their physical and chemical properties. It has been shown that in terms of quality and safety, all tested samples satisfied the requirements of the applicable documentation.

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1 Introduction

Packaging materials used in food industry are subject to rigid requirements and it should strongly comply the requirements of technical regulation TR TS 005/2011 "About safety of packaging" (Kisku & Swain, 2012; Schreiber et al. 2013). One of the mandatory conditions governing the use of packaging in food industry is the availability of a certificate of compliance issued by the Ministry of Health, which confirms packaging materials' safety for humans (Pantukhov, 2012a; Khan et al., 2014; Pantukhov, 2012a).

Currently, food industries and retailers who are widely using the packaging materials should not only ensure safety of the food products, but should also be economically viable and safe for human health and environment (Pantukhov 2012b). Therefore, various sanitary-epidemiological and hygienic requirements are applied to packaging products (Lu et al., 2010). Among this, some most common are (i) the packaging should not change organoleptic and physico-chemical properties of food products (ii) the packaging should not emit harmful or toxic substances (Lu et al., 2010; Liu, 2010; Rhim, 2013; Liu, 2015; Razavi, 2016), (iii) the packaging should not contain substances that have cumulative, mutagenic, allergenic and carcinogenic properties, and other compounds that affect the state and health of a living organism (Rhim, 2013) and (iv) the packaging should not be the nutritional medium for pathogenic and other microflora, and should not let through microorganisms.

This document established the requirements of the quality and composition of the material used for packaging, packaging surface, size and quality of side seams, strength and integrity of packaging (Liu, 2010, Kisku & Swain, 2012; Schreiber et al., 2013; Liu, 2015). This work is aimed to choose the optimum composition of biodegradable packaging materials based on gelatin and natural polysaccharides for obtaining films' samples and studying their properties.

2 Materials and methods

In present study gelatine and natural polysaccharides viz., agar-agar, carrageenan and hydroxypropylmethylcellulose (HPMC) has been used as packaging material. Based on the performance, physico-chemical properties and safety indicators: agar-agar (Helicon, USA), Kappa-carrageenan (Boc Sciences, USA), hydroxypropylmethylcellulose (Ashland Aqualon Functional Ingredients, USA), and gelatin ("GelatinesWeishardt", France) have been selected.

Gelatin is a colorless or yellowish partially hydrolyzed collagen protein with a transparent viscous mass, without taste and smell. It is a product of animals 'connective tissues processing

(denaturing). Further, gelatin couldn't dissolve in diluted acids and when it mixes with diluted acid it will swells. Swollen gelatin dissolves when heated and forming a solution which convert in jelly when solidifies (Rhim 2013).

Agar-agar is another gelatinous material which obtained from sea algae. Like gelatin it is also insoluble in cold water and can completely dissolve when heated at 85-95°C. Molecules of Carageenan are large, highly flexible, and can form inter twisted structures. It can form various gels at room temperature. Hydroxypropyl methyl cellulose is a stabilizer capable of maintaining and improving viscosity and consistency of food products (Rhim 2013).

Aiming to comprehensively study, various properties of the initial components for obtaining biodegradable polymers, their physico-chemical properties and indicators of chemical and microbiological safety were studied. During the study normative technical documentation for certain kinds of natural polysaccharides and gelatin were taken into account:

- GOST 16280-2002 - Food agar - Specifications (Schreiber et al., 2013)
- Carageenan. CAS 11114-20-8 (Schreiber et al., 2013)
- Hydroxypropylmethylcellulose - CAS 9004-65-3 (Schreiber et al., 2013)
- GOST 23058-89 - Gelatin as raw material for medical industry Specifications (Schreiber et al., 2013)

Mixture of Gelatin and other natural polysaccharides were aged in an autoclave in blowing mode for 15 minutes and films based on these were obtained by drying solutions of the tested components at various temperatures. The film-forming solution was transferred into a cuvette with (1-2) mm sides, which was covered with a glass cover. Then the solution was cooled at room temperature and one millimeter thick films were obtained by pouring. Film of the gelatin and other natural polysaccharides were dried up in various ways, among this some common are (i) drying at room temperature (ii) drying by air convection in a fume cabinet (iii) dried in a vacuum drying cabinet (the temperature of drying varied) and (iv) drying in a drying cabinet (the temperature of drying varied).

For obtaining transparent, flexible and uniform thickness films various combination of selected polysaccharide have been tried. As per the used combinations, film number such as No. 3 (mix of 20% agar and 80% carrageenan), No. 11 (a mixture of 30% agar and 70% carrageenan), No. 20 (a mixture of 40% agar and 60% carrageenan), No. 14 (a mixture of 20% hydroxypropylmethylcellulose and 80% gelatine), No. 16 (a mixture of 30% hydroxypropylmethylcellulose and 70% gelatin),

and No. 18 (a mixture of 40% hydroxypropylmethylcellulose and 60% gelatin) has been assigned. The mixture of components was kept on the heating mantle, the temperature of the mixer was 150 °C, the rotation speed was 800 rpm; the resulting solution was poured on a substrate with sides 2 mm thick; drying was performed in a drying cabinet at 85 °C.

3 Results

Date given in table 1 represented the physico-chemical properties of the selected and processed two agar-agar samples (sample A1 and A2). Classification of agar-agar was carried out on the basis of manufacturers. On the basis of physicochemical properties and technical basis, both the selected agar-agar samples fulfill the

requirement of GOST 16280-2002 for packaging materials. Results are in agreement with the findings and suggestions of Schreiber et al. (2013).

In physicochemical properties, agar-agar meets the criteria of GOST 16280-2002, in this study chemical and microbiological safety of agar-agar packaging material was also tested. Results of chemical and microbiological safety level of agar-agar have been represented in Table 2. Result of study revealed the presence of some heavy metals in agar-agar packaging material but contents of these heavy metals are not exceeding the maximum permissible concentration of GOST 16280-2002. During study presence of lead, cadmium, arsenic and mercury was reported. Presence of mesophilic aerobic and facultative anaerobic microorganisms was

Table 1 Physico-chemical properties of agar-agar

Indicator name	Sample		Requirements of GOST 16280-2002
	Sample A1	Sample A2	
Gel strength with mass fractions of 0.85% dry agar and 70% sugar (g)	1.650±165	1.600±160	Not less than 1.600
Loss of gel strength with the mass fraction of 0.85% dry agar after heating the solution for 2 h (%)	10.0±1.0	10.0±1.0	Not more than 10.0
Melting point of the gel with the mass fraction of 0.85% dry agar (°C)	82.0±8.2	85.0±8.5	Not less than 80.0
Gelatinization temperature of agar solution with the mass fraction of 0.85% dry agar (°C)	32.0±3.2	32.0±3.2	Not less than 30.0
Gelatinization temperature of agar solution with the mass fraction of 0.85% dry agar and 70% sugar (°C)	40.0±4.0	42.0±4.2	Not more than 42.0
Mass fraction of water (%)	16.0±1.6	17.5±1.8	Not more than 18.0
Mass fraction of ash (%)	4.0±0.4	4.2±0.4	Not more than 4.5
Presence of iodine	Not detected	Not detected	Not allowed
Mass fraction of substances insoluble in hot water (%)	0.20±0.02	0.30±0.03	Not more than 0.40

Here sample A1 – agar-agar made by Panreac (Germany); sample A2 – agar-agar made by Helicon (USA).

Table 2 Indicators of agar-agar chemical and microbiological safety

Indicator name	Sample		Requirements of GOST 16280-2002
	Sample A1	Sample A2	
Lead (mg/kg)	0.15±0.01	0.08±0.01	not more than 2.00
Cadmium (mg/kg)	below the determination limit	0.010±0.005	not more than 0.030
Arsenic (mg/kg)	0.010±0.005	0.010±0.005	no more than 0.500
Mercury (mg/kg)	below the determination limit	below the determination limit	not more than 0.02
Mesophilic aerobic and facultative anaerobic microorganisms (CFU/g)	0.9·10 ²	3.2·10 ²	not allowed
Coliform bacteria, Proteus group bacteria (CFU/g)	not found	not found	not allowed
Pathogens	not found	not found	not allowed

Here sample A1 – agar-agar made by Panreac (Germany); sample A2 – agar-agar made by Helicon (USA).

also reported during the study which is against the recommendation of GOST 16280-2002. Presence of coliforms, proteus group bacteria and pathogenic microorganisms was not reported from the agar-agar.

Results of physico-chemical properties of two carrageenan samples (K1 and K2) obtained from various manufacturers have been presented in Table 3. The results were analyzed in accordance with the requirements of the CODEX Alimentarius international standard for food products, CAS 11114-20-8. Result of study suggested that physico-chemical parameters of two selected carrageenan samples from various manufacturers did not exceed the values specified by CAS 11114-20-8. In this manner, in physicochemical characteristics, carageenan can be used as a packaging material.

Further, the results of testing carageenan chemical and

microbiological safety, including the contents of heavy metals and the microbiological parameters have been shown in Table 4. Data given in the table revealed that both the samples of carrageenan meet the safety requirements of CAS 11114-20-8 in physico-chemical and microbial properties. The contents of lead, cadmium, arsenic, mercury, mesophilic aerobic and facultative anaerobic microorganisms, coliforms, bacteria of group proteus, and pathogenic microorganisms did not exceed the maximum permissible concentration suggested by CsAS 11114-20-8.

Results of studied physicochemical parameters of two HPMC samples (samples G1 and G2) obtained from various manufacturers are shown in Table 5. Results were analyzed according to the requirements of the CODEX Alimentarius international standard (CAS 9004-65-3) for food products. The data in Table 5 shown that physico-chemical parameters two samples of HPMC did not exceed the values specified for packaging

Table 3 Physico-chemical characteristics of carageenan

Indicator name	Sample		Requirements of CAS 11114-20-8
	Sample K1	Sample K2	
Mass fraction of moisture (%)	10.5±1.0	11.0±1.1	Not more than 12.0
pH of the 1:100 suspension	9.5±1.0	10.0±1.0	8.0 – 11.0
Viscosity of a 1.5% solution at 75 °C (cPs)	6.5±0.3	5.5±0.3	Not less than 5.0
Mass fraction of sulfates (%)	25.0±2.5	18.0±1.8	15.0-40.0
Mass fraction of ash (%)	15.0±1.5	17.5±1.8	15.0-40.0
Mass fraction of ash insoluble in acid (%)	0.80±0.08	0.50±0.05	No more than 1.00
Mass fraction of substance insoluble in acid (%)	1.0±0.1	1.5±0.2	Not more than 2.0
Mass fraction of solvent, (%)	not found	0.050±0.005	Not more than 0.100

Note: sample K1 – Kappa-carageenan made by BocSciences (USA); sample K2 – iota-carageenan made by Newgreen Pharmchem Co. (China)

Table 4 Indicators of chemical and microbiological safety of carageenan

Indicator name	Sample		Requirements of CAS 11114-20-8
	Sample K1	Sample K2	
Lead (mg/Kg)	0.50±0.03	1.10±0.06	Not more than 5.0
Arsenic (mg/Kg)	0.20±0.01	0.50±0.03	Not more than 3.0
Cadmium (mg/Kg)	0.10±0.01	0.10±0.01	Not more than 2.0
Mercury (mg/Kg)	not found	not found	Not more than 1.0
MAFAM (CFU/g)	1.0·10 ²	1.5·10 ¹	Not more than 5.0·10 ³
Pathogens	not found	not found	Not allowed
Coliform bacteria (CFU/g)	not found	not found	Not allowed

Note: sample K1 – Kappa-carageenan made by BocSciences (USA); sample K2 – iota-carageenan made by Newgreen Pharmchem Co. (China)

Table 5 Physico-chemical characteristics of HPMC

Indicator name	Sample		Requirements of CAS 9004-65-3
	Sample G1	Sample G2	
Mass fraction of ash in terms of dry gelatin (%)	1.0±0.1	0.9±0.1	not more than 1.5
Decreased viscosity of the solution with the mass fraction of gelatin at 10% (%)	11.5±0.6	12.0±0.6	not more than 15.0
Dynamic viscosity of a solution with the mass fraction of gelatin of 10%, in terms of absolutely dry gelatin (MPa·s)	25.0±1.2	26.5±1.3	not less than 24.6
Strength of jelly with the mass fraction of gelatin of 10 %, in terms of absolutely dry ash-free gelatin (N gs)	14.7±0.7	15.0±0.8	Not less than 13.0
Transparency of the solution with the mass fraction of gelatin of 5 % (%)	75.0±3.8	80.0±4.0	Not less than 70.0
The melting point of the jelly with the mass fraction of gelatin of 10 %, in terms of absolutely dry ash-free gelatin (°C)	33.0±1.7	35.2±1.8	not less than 32.0
pH of the solution with mass share of gelatin of 1 %	5.5±0.5	5.7±0.6	5.2 – 6.1

Here, sample G1 – HPMC made by Acros (Belgium); sample G2 – HPMC made by Ashland Aqualon Functional Ingredients (USA)

Table 6 Indicators of chemical and microbiological safety of hydroxypropylmethylcellulose (HPMC)

Indicator name	Sample		Requirements of CAS 9004-65-3
	sample G1	sample G2	
Mass fraction of moisture (%)	8.0±0.8	8.5±0.9	Not more than 10.0
pH of the 1:100 solution	6.5	7.0	5.0 – 8.0
Mass fraction of sulfated ash (%)	1.2±0.1	1.5±0.1	Not more than 1.5 for samples with the viscosity of 50 cPs and higher. Not more than 3.0 for samples with the viscosity less than 50 cPs
Mesophilic aerobic and facultative anaerobic microorganisms (CFU/g)	not found	not found	Not allowed
Pathogens, including Salmonella	not found	not found	Not allowed
Coliform bacteria (CFU/g)	not found	not found	Not allowed

Here sample G1 – HPMC made by Acros (Belgium); sample G2 – HPMC made by Ashland Aqualon Functional Ingredients (USA)

materials CAS 9004-65-3. In case of viscosity, strength of jelly and transparency both used HPMS sample show superiority over the recommendation of CAS 9004-65-3.

Results of HPMC chemical and microbiological safety testing have been presented in table 6. Result of study revealed that both samples of HPMC meet the safety requirements of CAS 9004-65-3. The contents of lead, cadmium, arsenic, mercury, mesophilic aerobic and facultative anaerobic microorganisms, coliforms, bacteria of group proteus, and pathogenic microorganisms do not exceed the maximum permissible concentration. Among the tested two samples G1 (HPMC made by Acros, Belgium) have better properties as compared to sample G2 (HPMC made by Ashland Aqualon Functional Ingredients, USA) but no significant difference was found among these two.

The results of physico-chemical properties of gelatin (sample G1 and G2) have been given in Table 7. Both the used samples meet

the criteria specified by GOST 23058-89 in their physico-chemical parameters. Like HPMC, gelatin also performed better than the criteria prescribed by GOST 23058-89 and show good viscosity, strength of jelly and transparency. Further, gelatin have higher melting point which support it use as packaging material.

Chemical and microbiological safety of gelatin have been tested and reported that heavy metals, toxic elements, radionuclides, residual pesticides and microbiological parameters are in the range of GOST 23058-89 prescription (Table 8). The data in the table show that both samples of gelatin by their physico-chemical properties and safety indicators meet the requirements of GOST 23058-89. Further, contents of lead, cadmium, arsenic, mercury, mesophilic aerobic, facultative anaerobic microorganisms, coliforms, bacteria of group proteus, and pathogenic microorganisms are in the range of acceptance as it can use as packaging materials.

Table 7: Physico-chemical characteristics of gelatin

Indicator name	Sample		Requirements of GOST 23058-89
	Sample J1	Sample J2	
Duration of dissolution (min)	20.0±1.0	22.0±1.1	not more than 25.0
Mass fraction of moisture (%)	12.9±0.5	13.0±0.5	not more than 16.0
Mass fraction of ash in terms of absolutely dry gelatin (%)	1.0±0.1	0.9±0.1	not more than 1.5
Decreased viscosity of the solution with the mass fraction of gelatin of 10% (%)	11.5±0.6	12.0±0.6	not more than 15.0
Dynamic viscosity of a solution with the mass fraction of gelatin of 10%, in terms of absolutely dry gelatin (MPa·s)	25.0±1.2	26.5±1.3	not less than 24.6
Strength of jelly with the mass fraction of gelatin of 10%, in terms of absolutely dry ash-free gelatin, N (gs)	14.7±0.7	15.0±0.8	Not less than 13.0
Transparency of the solution with the mass fraction of gelatin of 5% (%)	75.0±3.8	80.0±4.0	Not less than 70.0
The melting point of the jelly with the mass fraction of gelatin of 10%, in terms of absolutely dry ash-free gelatin (°C)	33.0±1.7	35.2±1.8	not less than 32.0
pH of the solution with mass share of gelatin of 1%	5.5±0.5	5.7±0.6	5.2 – 6.1

Here sample J1 – gelatin made by "Minvody Gelatin Plant" (Russia); sample J2 – gelatin made by "GelatinesWeishardt" (France)

Table 8 Indicators of chemical and microbiological safety of gelatin

Indicator name	Sample		Requirements of GOST 23058-89
	Sample J1	Sample J2	
Sulphurous acid (in equivalent to SO ₂ , %)	0.010±0.001	below the determination limit	no more than 0.060
Zinc (mg/kg)	10.00±0.50	20.00±1.00	not more than 100.00
Lead (mg/kg)	0.15±0.01	0.08±0.01	not more than 2.00
Cadmium (mg/kg)	below the determination limit	0.010±0.005	not more than 0.030
Copper (mg/kg)	below the determination limit	below the determination limit	not more than 15.00
Arsenic (mg/kg)	0.010±0.005	0.010±0.005	no more than 0.500
Mercury (mg/kg)	below the determination limit	below the determination limit	not more than 0.02
Mesophilic aerobic and facultative anaerobic microorganisms (cells in 1 g of gelatin)	0.9·10 ²	3.2·10 ²	not more than 1.0·10 ³
Coliform bacteria (cells per 1 g of gelatin)	not found	not found	not allowed
Protea group bacteria	not found	not found	not allowed
Gelatin containing bacteria (cells per 1 g of gelatin)	not found	not found	not allowed
Pathogens	not found	not found	not allowed

Here sample J1 – gelatin made by "Minvody Gelatin Plant" (Russia); sample J2 – gelatin made by "GelatinesWeishardt" (France)

Gelatine and natural polysaccharides were used in various concentration and total 21 combination were prepared which were assigned sample number 1-21 (table 9). All the predescribed packaging material combination were mixed in required amount of distilled water and heat upto dissolution. After dissolution, the mixture of components was kept on the heating mantle, the

temperature of the mixer was 150 °C, the rotation speed was 800 rpm; the resulting solution was poured on a substrate with sides 2 mm thick; drying was performed in a drying cabinet at 85 °C (Table 9).

Result of drying gelatin-based solutions and natural polysaccharides

Table 9 Composition of studied gelatin-based films and natural polysaccharides

Sample number	Amount of the ingredient (wt. %)				
	Carageenan	HPMC	Agar-agar	Gelatin	Water
1	5.0	2.5	–	5.0	87.5
2	10.0	2.5	–	10.0	77.5
3	5.0	5.0	–	51.0	39.0
4	10.0	5.0	–	5.0	80.0
5	5.0	10.0	–	30.0	55.0
6	10.0	10.0	–	51.0	29.0
7	5.0	–	2.5	5.0	87.5
8	10.0	–	2.5	10.0	77.5
9	5.0	–	5.0	30.0	60.0
10	10.0	–	5.0	51.0	34.0
11	5.0	–	10.0	51.0	34.0
12	10.0	–	10.0	10.0	70.0
13	–	2.5	2.5	5.0	90.0
14	–	10.0	2.5	10.0	77.5
15	–	5.0	10.0	10.0	75.0
16	5.0	2.5	10.0	51.0	31.5
17	10.0	2.5	2.5	10.0	75.0
18	5.0	5.0	5.0	5.0	80.0
19	10.0	5.0	5.0	10.0	70.0
20	5.0	10.0	10.0	30.0	45.0
21	10.0	10.0	10.0	10.0	60.0

Table 10 Results of drying gelatin-based solutions and natural polysaccharides

Sample number	Result of drying
1	The film was not obtained due to low components' concentrations
2	High-quality film could not be obtained; there are undissolved HPMC particles
3	Transparent and elastic films of uniform thickness
4	High-quality film could not be obtained; there are undissolved HPMC particles
5	The obtained samples are fragile and have brittle structure. It is inappropriate to use them for further research
6	The obtained samples are fragile and have brittle structure. It is inappropriate to use them for further research
7	The obtained samples are fragile and have brittle structure. It is inappropriate to use them for further research

Sample number	Result of drying
8	The obtained films have uneven thickness due to high viscosity of the solution. It is inappropriate to use them for further research
9	The obtained samples are fragile and have brittle structure. It is inappropriate to use them for further research
10	The obtained films have uneven thickness due to high viscosity of the solution. It is inappropriate to use them for further research
11	Transparent and elastic films of uniform thickness
12	The obtained films have uneven thickness due to high viscosity of the solution. It is inappropriate to use them for further research
13	Plasticity and flexibility of the films are missing, therefore, it is impractical to use them for further research
14	Transparent and elastic films of uniform thickness
15	Plasticity and flexibility of the films are missing, therefore, it is impractical to use them for further research
16	Transparent and elastic films of uniform thickness
17	The obtained films have uneven thickness due to high viscosity of the solution. It is inappropriate to use them for further research
18	Transparent and elastic films of uniform thickness
19	The obtained films have uneven thickness due to high viscosity of the solution. It is inappropriate to use them for further research
20	Transparent and elastic films of uniform thickness
21	Plasticity and flexibility of the films are missing, therefore, it is impractical to use them for further research

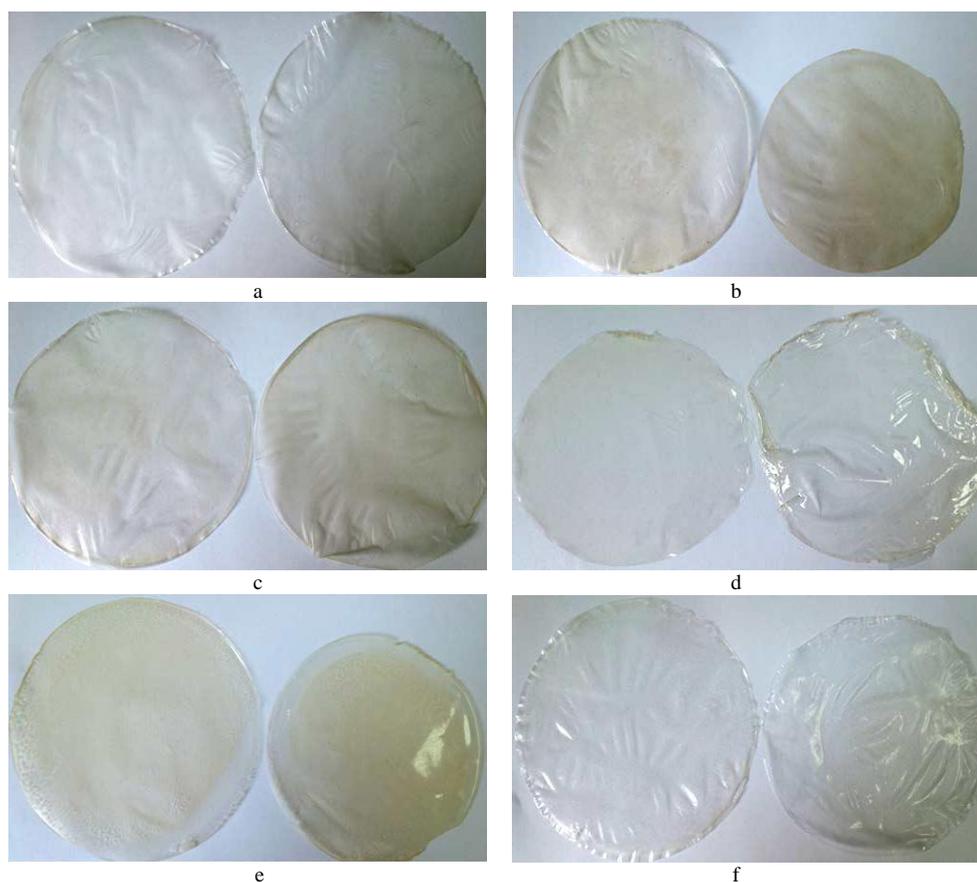


Figure 1 Photos of the studied films sample No. 3 (a); sample No. 11 (b); sample No. 14 (c); sample No. 16 (d); sample No. 18 (e) and sample No. 20 (f)

Table 11 Characteristics of gelatin and natural polysaccharides based packaging materials obtained using various technological methods

Indicator Name	Value of the indicator for packaging materials obtained by various processing methods		
	Extrusionblowing	thermoforming	extrusion blowing forming
Sample No. 3			
Tensile strain σ_p , MPa	56.8±5.7	15.0±1.5	32.6±3.3
Strain-to-failure, ϵ_p , %	10.0±1.0	1.2±0.1	5.4±0.5
Elasticity modulus, MPa	2,550±255	550±55	1,250±125
Gas permeability, $\text{cm}^3/\text{m}^2 \cdot 24\text{h} \cdot \text{atm}$	246.000±24.600	5.665±0.567	75.006±7.501
Sample No. 11			
Tensile strain σ_p , MPa	88.3±8.8	13.8±1.4	25.5±2.6
Strain-to-failure, ϵ_p , %	7.7±0.8	2.0±0.2	6.0±0.6
Elasticity modulus, MPa	1,800±180	400±40	980±98
Gas permeability, $\text{cm}^3/\text{m}^2 \cdot 24\text{h} \cdot \text{atm}$	310.002±31.000	8.124±0.812	115.225±11.523
Sample No. 14			
Tensile strain σ_p , MPa	63.7±6.4	11.9±1.2	22.1±2.2
Strain-to-failure, ϵ_p , %	9.5±1.0	1.5±0.1	3.9±0.4
Elasticity modulus, MPa	3,300±330	780±78	1,100±110
Gas permeability, $\text{cm}^3/\text{m}^2 \cdot 24\text{h} \cdot \text{atm}$	285.015±28.502	6.075±0.608	80.905±8.091
Sample No. 16			
Tensile strain σ_p , MPa	72.5±7.3	9.5±1.0	34.0±3.4
Strain-to-failure, ϵ_p , %	7.5±0.8	1.0±0.1	4.4±0.4
Elasticity modulus, MPa	3100±310	390±39	1500±150
Gas permeability, $\text{cm}^3/\text{m}^2 \cdot 24\text{h} \cdot \text{atm}$	324,005±32,401	8,125±0,813	111,325±11,133
Sample No. 18			
Tensile strain σ_p , MPa	67.0±6.7	14.4±1.4	37.2±3.7
Strain-to-failure, ϵ_p , %	11.1±1.1	2.8±0.3	5.2±0.5
Elasticity modulus, MPa	2700±270	520±52	675±68
Gas permeability, $\text{cm}^3/\text{m}^2 \cdot 24\text{h} \cdot \text{atm}$	415,007±41,501	10,122±1,012	100,045±10,005
Sample No. 20			
Tensile strain σ_p , MPa	72.0±7.2	10.2±1.0	28.4±2.8
Strain-to-failure, ϵ_p , %	8.8±0.9	1.9±0.2	6.1±0.6
Elasticity modulus, MPa	1450±145	645±65	800±80
Gas permeability, $\text{cm}^3/\text{m}^2 \cdot 24\text{h} \cdot \text{atm}$	500,009±50,001	9,005±0,900	85,165±8,517

film formulation have been presented in table 10. As per the used combinations, only six samples of gelatin-based films and natural polysaccharides (3, 11, 14, 16, 18 and 20) can formed transparent and elastic film with uniform thickness. Above said combination used for further study while rest of the combination were discard

from the further study (Figure 1, Table 10).

The results of determining structural and mechanical properties along with strength and gas permeability of the obtained samples (3, 11, 14, 16, 18 & 20) have been given in Table 11.

4 Discussion

Packaging is an important element for preserving food quality. According to the normative documentation, a developer of new products should take care of consumer properties and safety, their hygienic standards, requirements for ensuring compliance with specified regulations in the process of production, storage, transportation and sales. In addition, packaging material that contacts food products should have certain adaptability to manufacturing and use, as well as fuel efficiency, reliability, strength, attractiveness and, most importantly, be comfortable for the consumer.

To choose the optimal composition of biodegradable polymers, the following samples of gelatin and natural polysaccharides were chosen based on the studies for determining their physico-chemical properties and safety: agar (Helicon, USA), Kappa-carrageenan (Boc Sciences, USA), hydroxypropylmethylcellulose (Ashland Aqualon Functional Ingredients, USA), gelatin ("Gelatines Weishardt", France). By the indicators of chemical and microbiological safety, all studied samples of gelatin comply with the current hygienic standards. Result of present study are in agreement with the finding and suggestion of Lu et al. (2010); Liu (2010); Rhim (2012); Liu (2015) and Razavi (2016).

In this regard, for further development of the technology of obtaining biodegradable polymers, it is advisable to use any presented samples of natural polysaccharides and gelatin as the components.

Conclusion

Physico-chemical properties and safety indicators of gelatin and natural polysaccharides (agar, carageenan, hydroxypropylmethylcellulose – HPMC) have been studied. It has been shown that in terms of quality and safety, all tested samples satisfy the requirements of the applicable documentation. With the aim of finding the optimal composition of biodegradable packaging materials based on gelatin and natural polysaccharides, film samples have been obtained.

Conflict of Interest

Authors would hereby like to declare that there is no conflict of interests that could possibly arise

References

- Khan A, Huq T, Khan RA, Riedl B, Lacroix M (2014). Nanocellulose-Based Composites and Bioactive Agents for Food Packaging. *Critical Reviews in Food Science and Nutrition* 54: 163–174.
- Kisku SK, Swain SK (2012) Study of oxygen permeability and flame retardancy properties of biodegradable polymethylmethacrylate. *Polymer Composites* 33: 79–84.
- Liu M (2010) Analysis of biodegradability of biodegradable mulching films. *Journal of Polymers and the Environment* 18: 148–154.
- Liu SJ (2015) Rheological Properties and Scaling Laws of kappa-Carrageenan in Aqueous Solution. *Macromolecules* 20: 7649-7657.
- Lu G, Peng X, Sun L, Wang P, Zhu G (2010) Patent 101812188A China, MPK 08 K 13/06. Method for preparing starch-based bio-resin suitable for blowing films; the applicant and the patentee Gangdongshangjiu biodegradable plastics CO LTD. No.CN201019050014 20100202; stat. 02.02.2010; publish. 2.02.2010.
- Pantuykhov PV (2012a) Interaction of natural fillers with polyethylene matrix in biodegradable polymer compositions. Proceedings of the XII Annual International Conference of Biochemical Physics RAS-Universities "Biochemical Physics" held on 13-15 January, 2012 at Moscow, Pp. 131–133.
- Pantuykhov PV (2012b) The oxidative and biological destruction of composite materials based on low-density polyethylene and lignocellulosic fillers. *Chemistry and Chemical Technology* 6:349–354.
- Razavi SMA (2016) Structural and physicochemical characteristics of a novel water-soluble gum from Lallemandiaroyleana seed. *International Journal of Biological Macromolecules*, 83: 142–151.
- Rhim JW (2013) Bio-nanocomposites for food packaging applications. *Progress in Polymer Science* 38: 1629–1652.
- Schreiber SB, Bozell JJ, Hayes DG, Zivanovic SV (2013) Introduction of primary antioxidant activity to chitosan for application as a multifunctional food packaging material. *Food Hydrocolloids* 33: 207–214.